#### de novo sequencing of the sunflower genome



#### **Stéphane Muños**

LIPM – INRA Toulouse

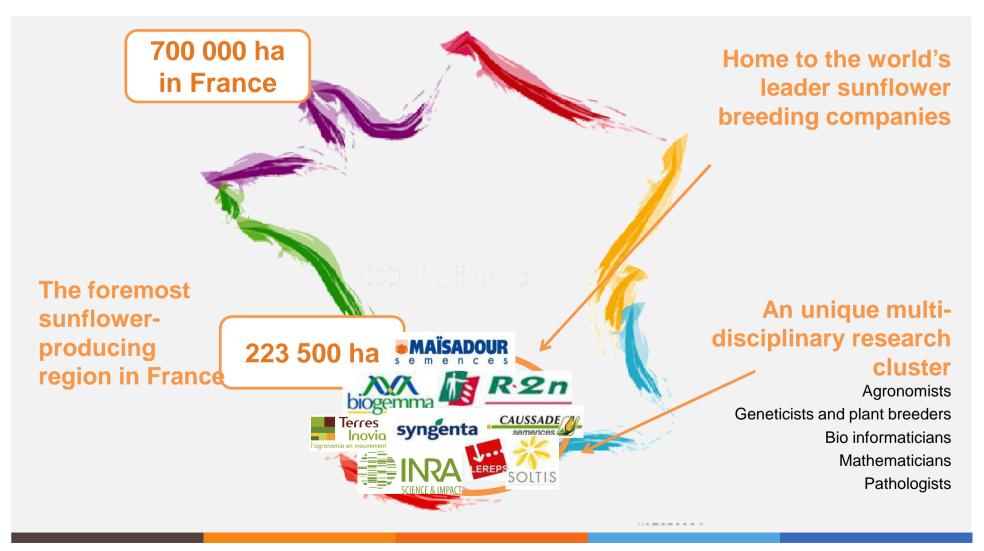
stephane.munos@toulouse.inra.fr @stephane\_munos @SUNRISE\_France

## Sunflower, an important crop for Europe **Million hectares 39** Million tons of seed produced worldwide 30 worldwilde 71% 80% in Europe in Europe The global production of sunflower seeds has to increase Societal challenge to meet growing demand (human food, animal feed, green chemistry...)

, ,



# The french region Midi-pyrénées, a key player in global sunflower production





## Sequencing of the sunflower genome

Jérôme Gouzy, Baptiste Mayjonade, Christopher J. Grassa, Sébastien Carrère, Erika Sallet, Ludovic Legrand, Hélène Badouin, Nicolas Pouilly, Marie-Claude Boniface, Nicolas Blanchet, Brigitte Mangin, Cécile Donnadieu, Hélène Bergès, Stéphane Muños, Patrick Vincourt, Nicolas Langlade

Christopher J. Grassa, Navdeep Gill, Thuy Nguyen, Nolan Kane, Loren H. Rieseberg

John E. Bowers, John M. Burke









N. Langlade

UBC Vancouver L. Rieseberg

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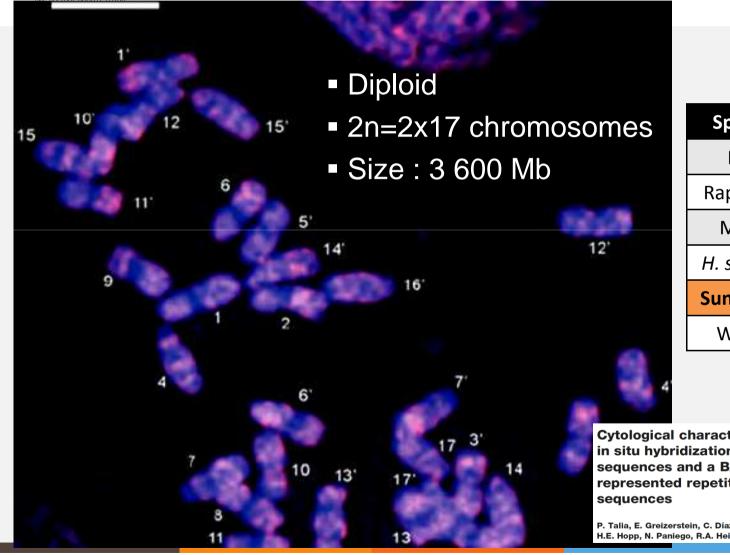








## Sunflower genome background



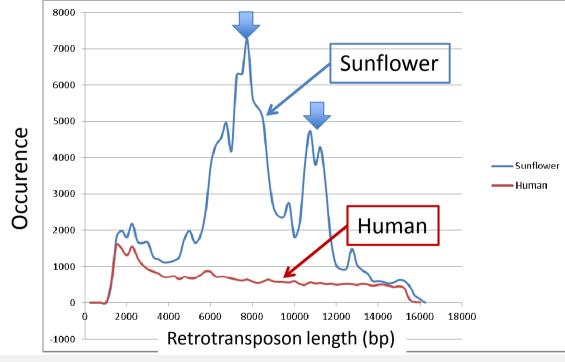
Size
430 Mb
1 100 Mb
2 300 Mb
3 200 Mb
3 600 Mb
17 000 Mb

Cytological characterization of sunflower by in situ hybridization using homologous rDNA sequences and a BAC clone containing highly represented repetitive retrotransposon-like sequences

P. Talia, E. Greizerstein, C. Díaz Quijano, L. Peluffo, L. Fernández, P. Fernández, H.E. Hopp, N. Paniego, R.A. Heinz, and L. Poggio



Length distribution of LTR retrotransposons



LTRharvest (Ellinghaus et al. 2008, default parameters)



Repeats = 33% of the sunflower genome

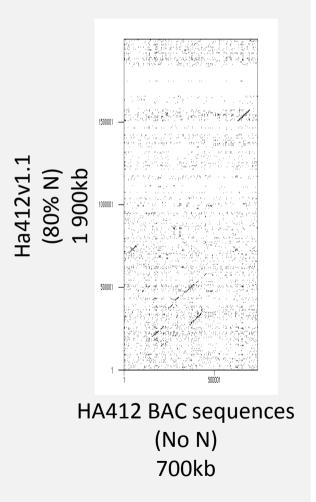
Repeats = 8% of the Human genome

Two major repeats in the sunflower genome: 8kb and 11.5kb

The repeats make the assembling very difficult



- Sunflower line : HA412
   International Consortium
   UBC Vancouver, INRA Toulouse, UGA Athens
  - Produced from 454 and Illumina sequencing
  - 1 989 Mb (55% of 3.6Gb)
  - Genome browser and annotation on
     <u>www.heliagene.org</u>
  - Good at macro scale but local assembly problems





•Sunflower line : HA412

International Consortium



UBC Vancouver, INRA Toulouse, UGA Athens

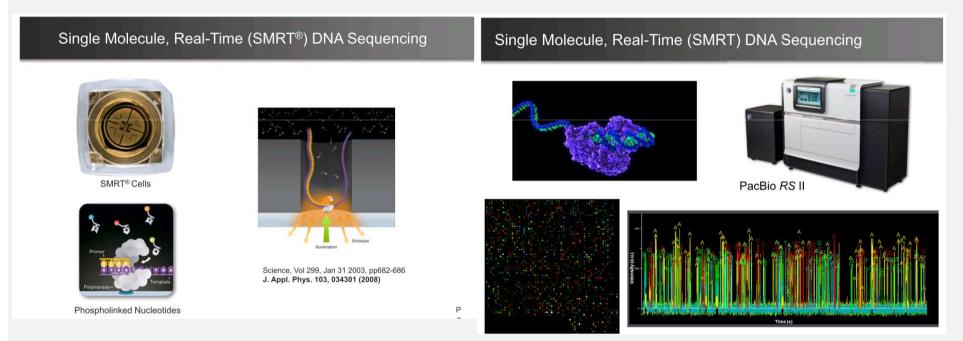
A lot of genetic and genomic ressources produced: BAC libraries, physical map, high-density genetic maps, SNPs from re-sequenced genomes...

Difficulties were due to the short length of the sequences used for assembly that cannot span the long repeats



# At the end of 2014 : a technological breakthrough

PacBio RSII (Pacific Biosciences)

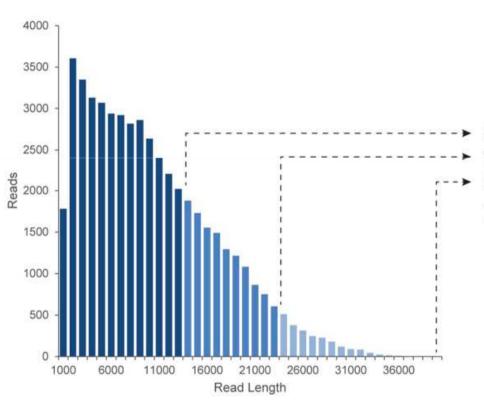


A movie (fluorescence incorporation during the synthesis) is recorded and then converted to DNA sequence



# At the end of 2014 : a technological breakthrough

#### P6-C4: Read Length Performance



Half of data in reads:	> 14 kb	
Top 5% of reads:	> 24 kb	
Maximum read length:	> 40 kb	
Data per SMRT <sup>®</sup> Cell:	500 Mb – 1 Gb	(in 4 hours)

## PacBio produces sequences longer than the known repeats

P6-C4, 4-hr movie, 20-kb BluePippin™ size-selected E. coli library (1 SMRT Cell)



SUNRISE Project (2012-2019) INRA Toulouse (LIPM, CNRGV, Genomic Platform) First PacBio in France (GeT-PlaGe)

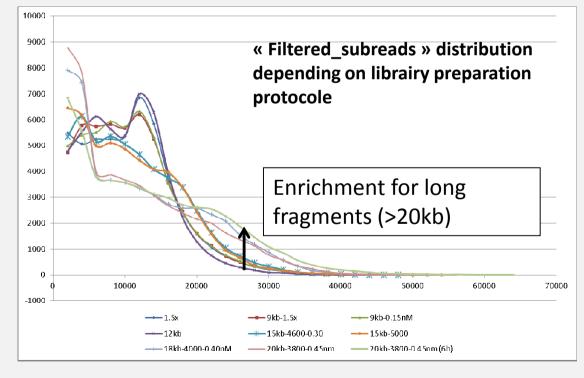
### **Sunflower line: XRQ**

#### 100% PacBio data used for the assembling.



#### The longer the PacBio sequences are, the better it is to span LTR :

- New DNA extraction protocole (submitted to BioTechniques)
- Optimisation of fragmentation, purification, loading
- Increase run time to  $4 \rightarrow 6h$  (movie-length)



## Extraction of high-molecular-weight genomic DNA for long-read sequencing of single molecules

Baptiste Mayjonade<sup>1</sup>, Jérôme Gouzy<sup>1</sup>, Cécile Donnadieu<sup>2</sup>, Nicolas Pouilly<sup>1</sup>, William Marande<sup>3</sup>, Caroline Callot<sup>4</sup>, Nicolas Langlade<sup>1</sup>, and Stéphane Muños<sup>1</sup>

<sup>1</sup>LIPM, Université de Toulouse, INRA, CNRS, Castanet-Tolosan, France, <sup>2</sup>Get-PLAGE, Université de Toulouse, INRA, CNRS, Castanet Tolosan, France, <sup>3</sup>CNRGV, Université de Toulouse, INRA, CNRS, Castanet Tolosan, France, and <sup>4</sup>CRCT, INSERM, Université de Toulouse, CNRS, Toulouse, France

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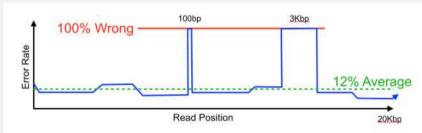
B. Mayjonade





#### 1) Correction of the raw sequences

J. Gouzy



https://dazzlerblog.wordpress.com/2015/11/06/intrinsic-quality-values/

1.1: Pairwise comparison of raw sequences (PBCR/MHAP, minimap, falcon)

1.2: The sequences are corrected

#### 2) Assembling of the corrected sequences

2.1: Pairwise comparison of the corrected sequences (WGS/Falcon)

2.2: Sequences are aligned (based on the overlap between sequences), the contig is breaked on the point where the repeat is not spanned.

2.3: consensus sequences of the contigs

#### 3) « Polishing » of the consensus contigs sequences

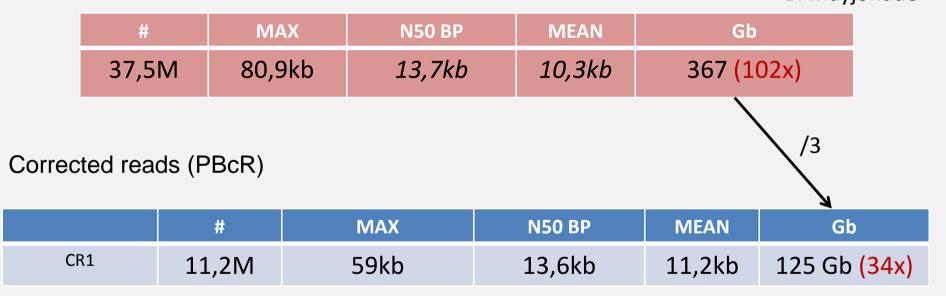
3.1: mapping of the raw data on the consensus sequences (Blasr)

3.2: correction of the consensus sequence based on the error rate model of the polymerase (quiver)



Raw data (407 SMRT cells)

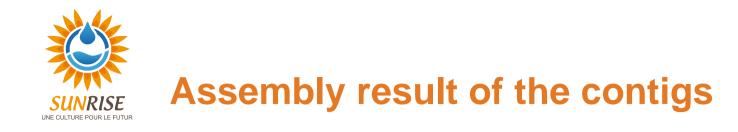




Third of the raw data are conserved after correction and used for assembling







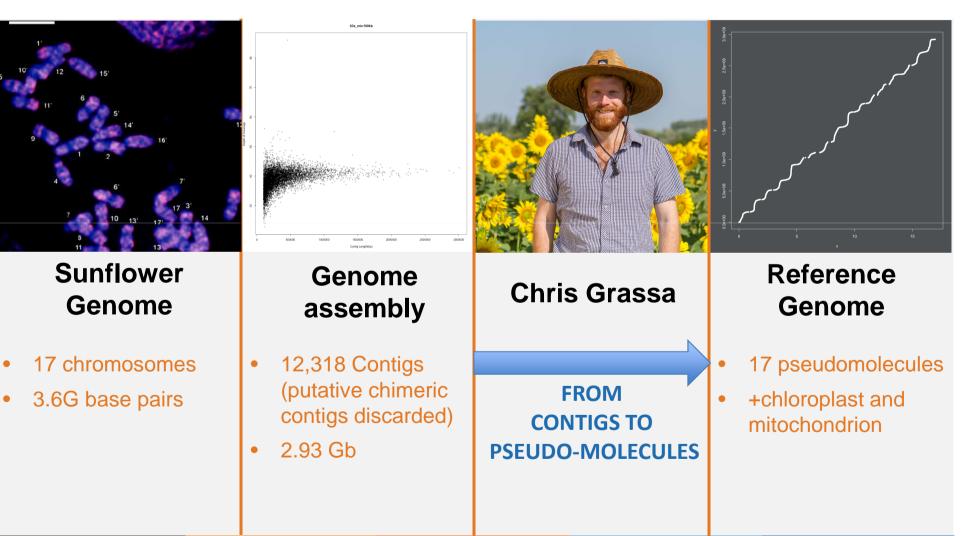
#ctg	ΜΑΧ	N50 BP	# > N50	MEDIAN	Gb
13 124	4.4M	498 kb	1700	118 kb	3.03

## →80% of genome in the contigs (No Ns)

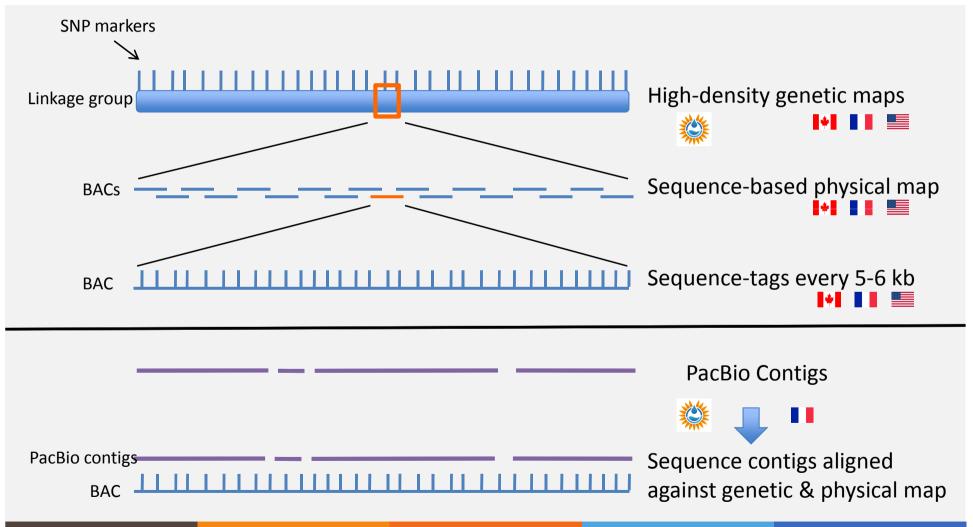
→20% not assembled (likely concatemeres of rDNA, TE, telomeres, centromeres)



## From contigs to chromosome sequences





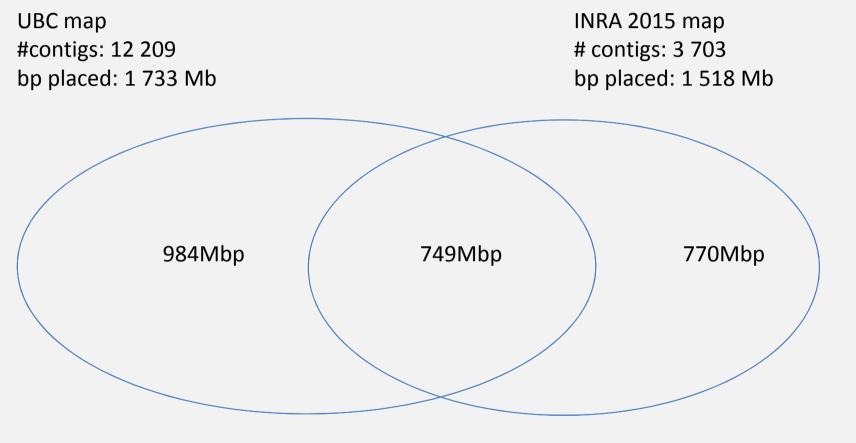




## **Integration of High-Density Genetic Maps**

INRA (Muños)	UGA (Bowers)	<b>USDA</b> (Talduker)	<b>UBC</b> (Grassa)
• 86,223 markers	• 10,080 markers	<ul> <li>5,019 RAD-tag markers</li> </ul>	<ul> <li>Sequenced-based (~2.5M SNPs)</li> </ul>
<ul> <li>3 Populations:         <ul> <li>HA89 x LR1</li> <li>XRQ x PSC8 - 2014</li> <li>XRQ x PSC8 - 2015</li> </ul> </li> </ul>	<ul> <li>4 Populations:         <ul> <li>HA412 x RHA415</li> <li>HA412 x ANN1238</li> <li>NMS373 x Hopi</li> <li>RHA280 x RHA801</li> </ul> </li> </ul>	<ul> <li>3 F2 Populations:         <ul> <li>HA89 x RHA464</li> <li>B-line x RHA464</li> <li>CR29 x RHA468</li> </ul> </li> </ul>	• <b>1 Population:</b> - RHA280 x RHA801



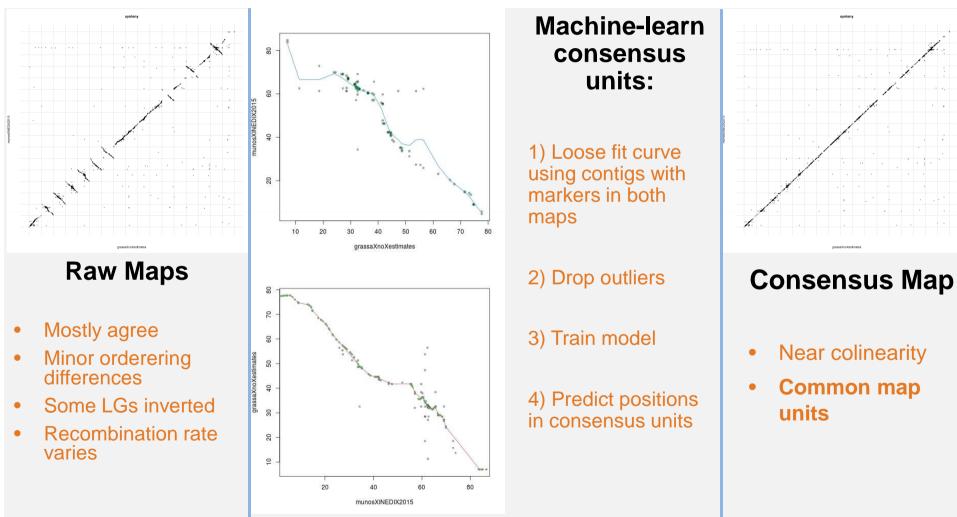


Other maps (INRA 2014, UGA, USDA): 415Mbp

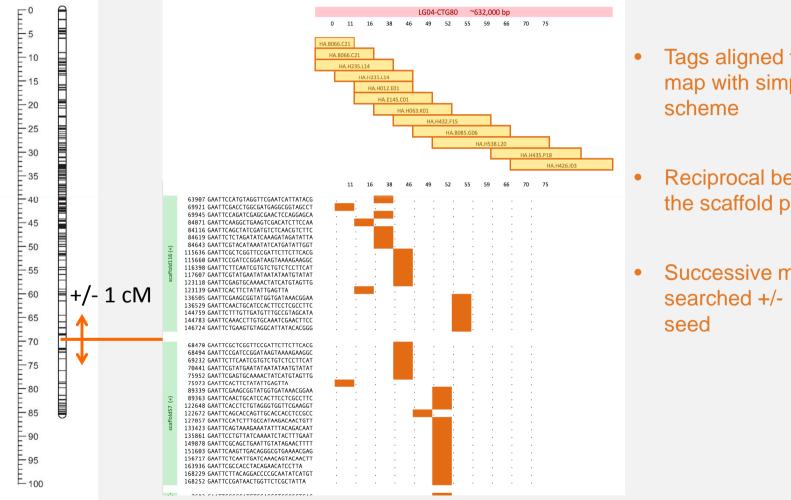


## **Build Consensus Map Units**

C. Grassa

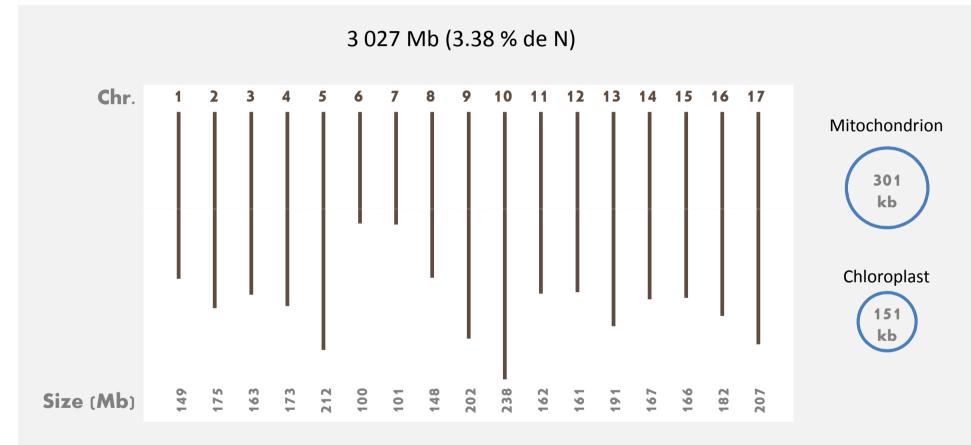






- Tags aligned to physical map with simple scoring
- Reciprocal best hits seed the scaffold position
- Successive matches searched +/- 1cM from





98.5% of contigs in the pseudomolecules



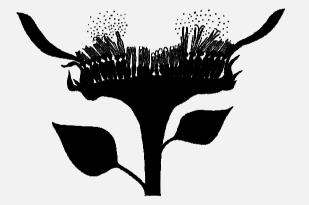
61 RNA-Seq librairies on the sequenced genotype (XRQ)

Organ-specific expression (12 organs)

Abiotic stress response: drought, osmotic stress, salt stress (in roots and leaves) Hormone regulation: (9 hormones in roots and leaves)

#### **Gene annotation**

52 243 protein coding genes (mRNA)
4 945 IncRNA genes
88 pre-miRNA genes (351 mature miRNA)
862 tRNA and rRNA genes

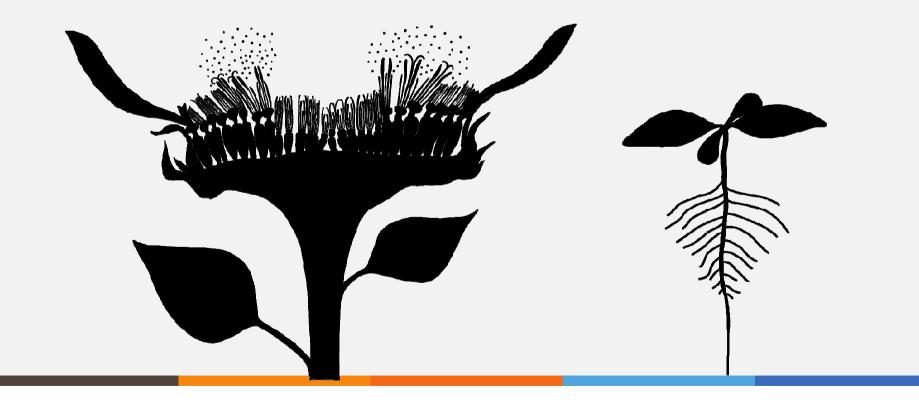


#### 98% of transcripts mapped on pseudo-molecules



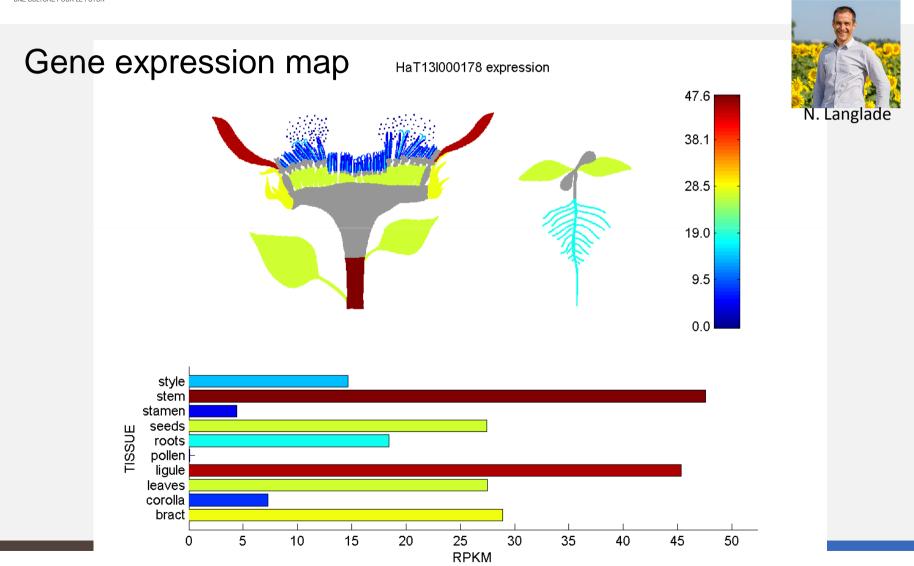
#### Expression data

Maintain access to raw count data Integrate visualization on <u>www.heliagene.org</u>



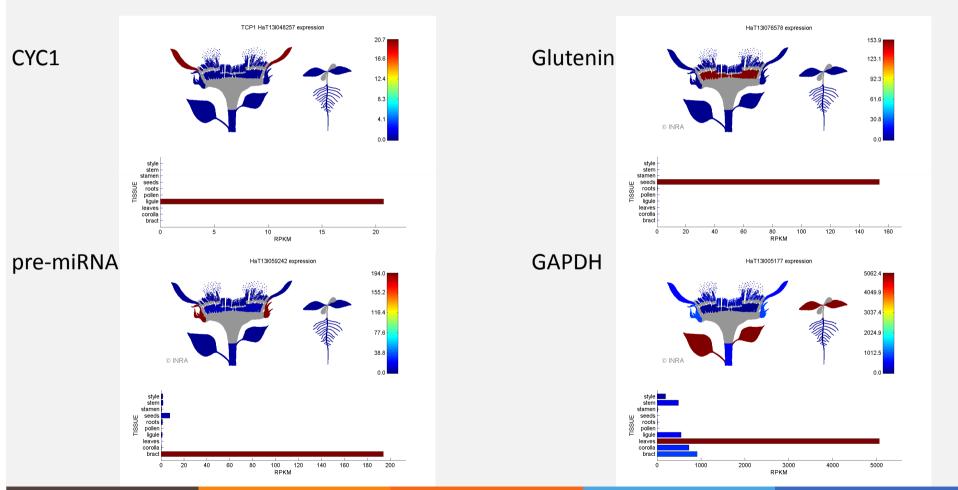


## **User interface and data visualization**





#### **Examples of organ-specific genes**





## What next for sunflower?

# Improving the sunflower assembly.

#### **Optical mapping**

CNRGV-INRA Toulouse (N. Rodde, C. Chantry, H. Bergès) Irys system (Bionano) acquired in March 2016

**NRGene on HA412 line** 



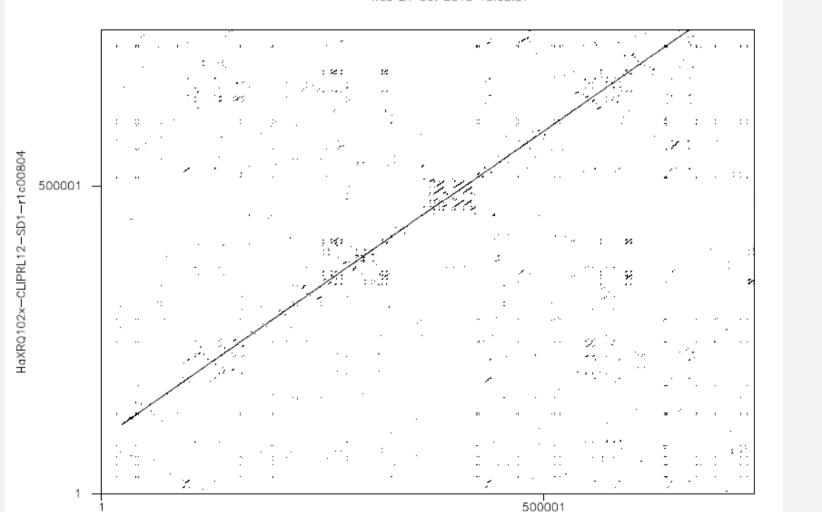
## What Next for sunflower?

## Several reference genomes are needed because, nor XRQ nor HA412 represent the averall genetic variations in sunflower

A first step : PSC8 sunflower line de novo sequenced (50X PacBio data, HeliOr project)

#ctg	MAX	N50 BP	# > N50	MEDIAN	Gb
26 273	2.5M	223kb	3799	66 kb	3.15





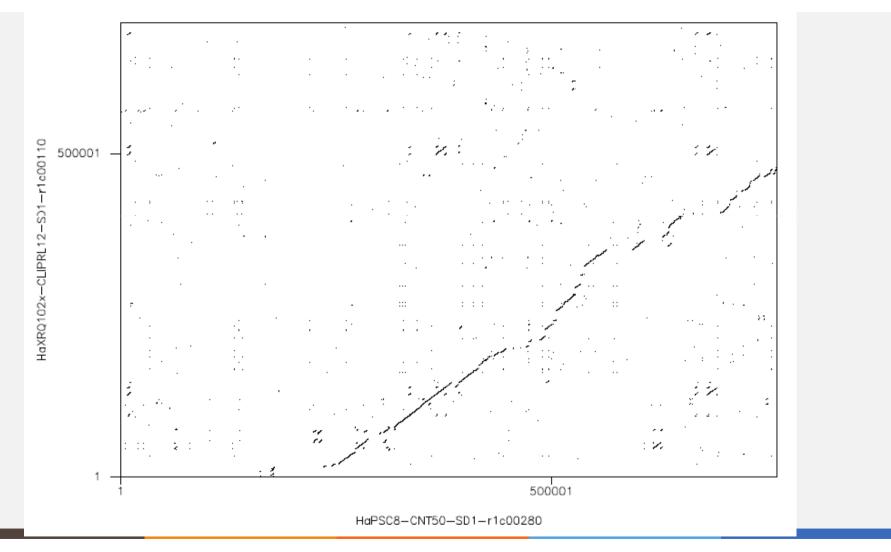
Wed 21 Oct 2015 13:02:37

HaPSC8-CNT50-SD1-r1c00306



## **XRQ vs PSC8: regions with structural variations**

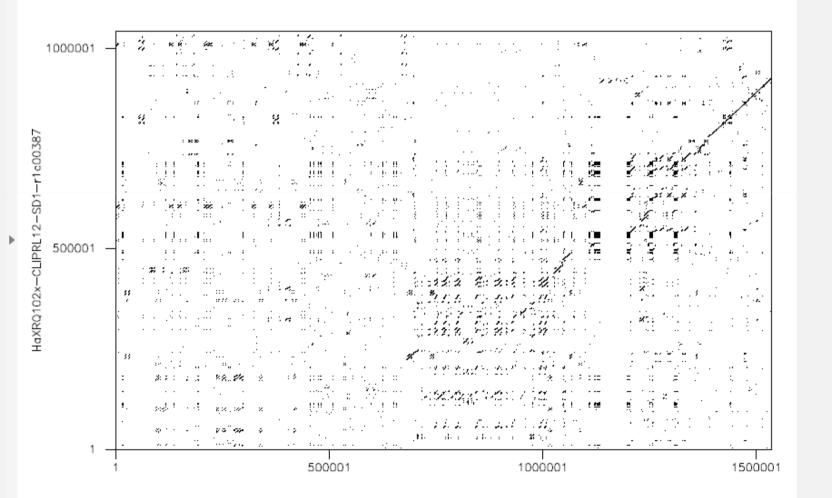
Mon 19 Oct 2015 11:07:44





#### XRQ vs PSC8: highly divergent regions

Wed 21 Oct 2015 13:06:52



HaPSC8-CNT50-SD1-r1c00033



#### A high quality genome sequence produced (XRQ line)

www.heliagene.org

## Sunflower could be a model plant like tomato became for fleshy fruits!

But breeding and genetic research need more genomes to be sequenced and more tools and data.



#### Felicity Vear (INRA Clermont-Ferrand)



#### Patrick Vincourt (INRA Toulouse)



## Thank you for your attention





Partners

syngenta biogemma and the series of southers and southers