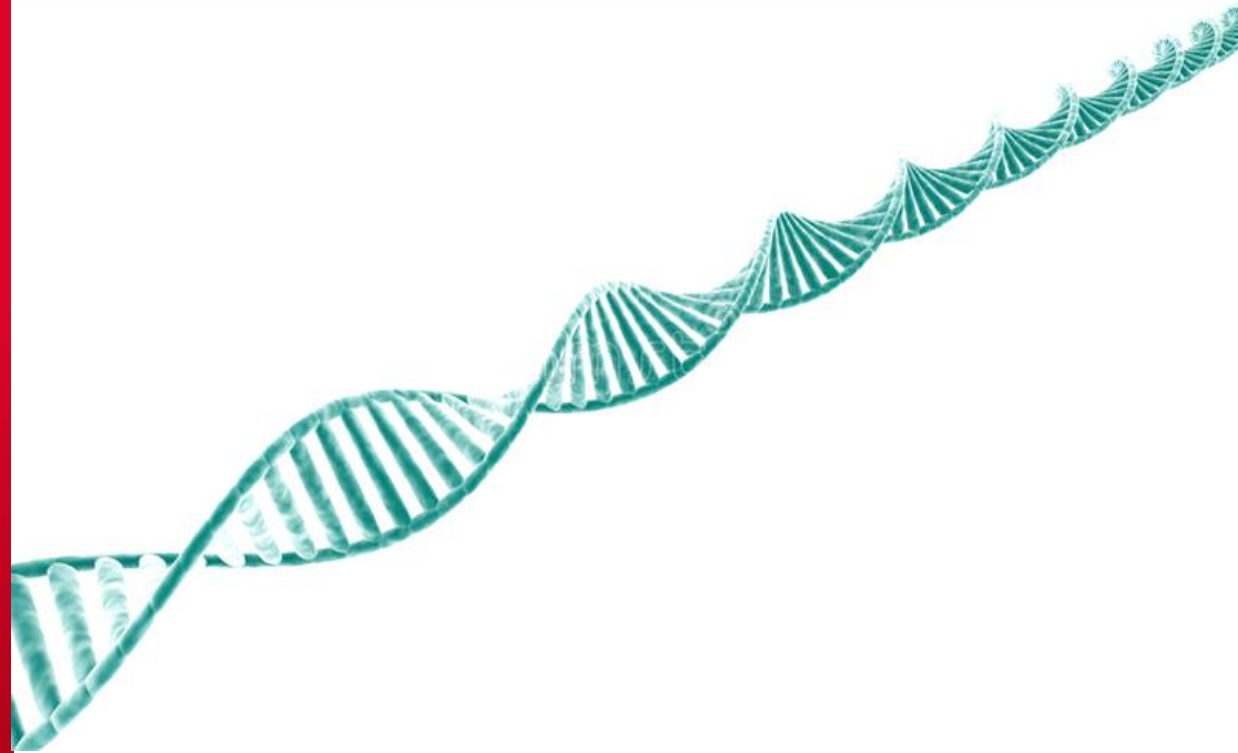


DE LA RECHERCHE À L'INDUSTRIE

cea



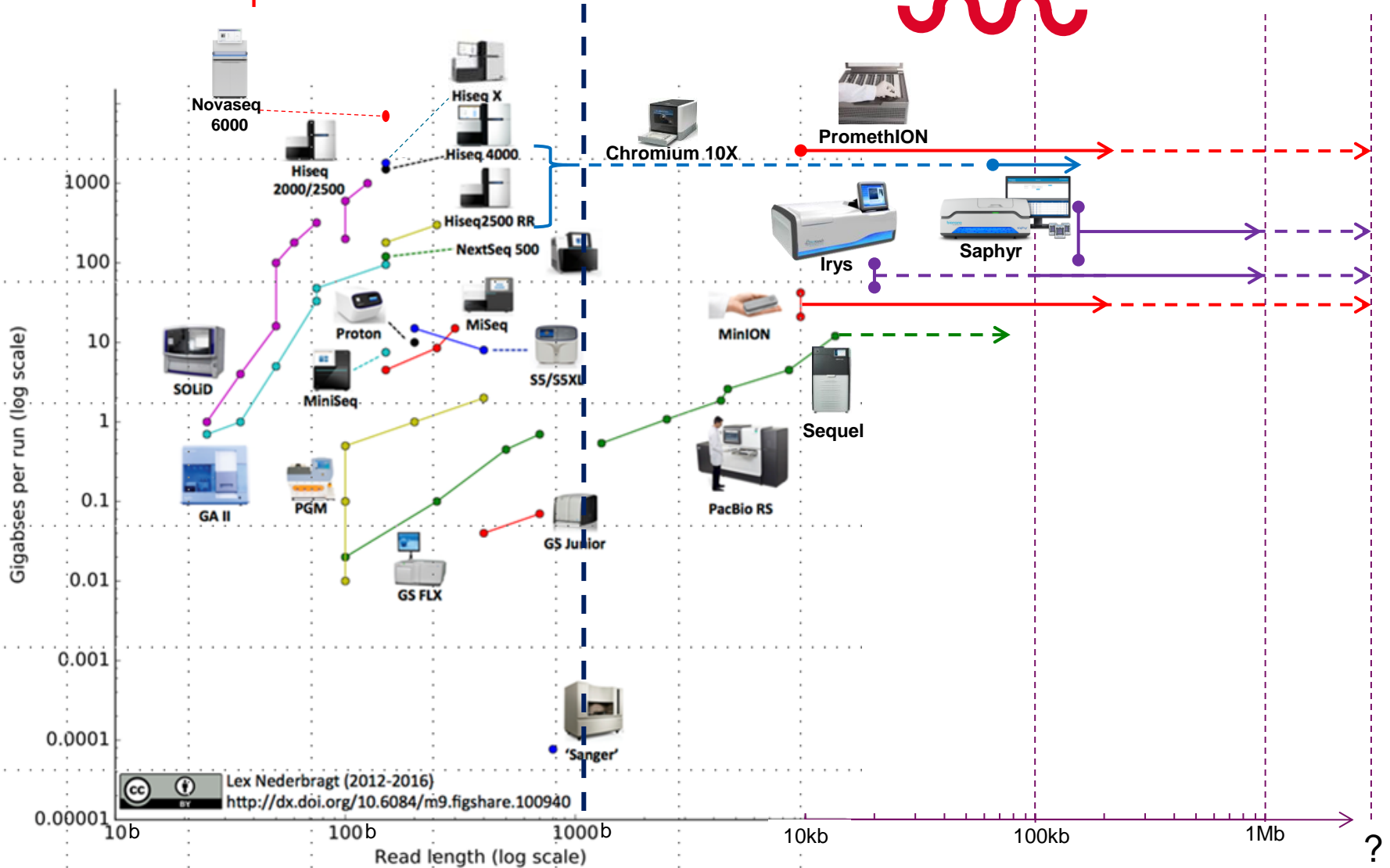
Assemblage *de novo* avec les technologies
ONT et Bionano Genomics:
l'exemple *Brassica*

Erwan DENIS et Cyril FALENTIN

Colloque EPGV 2018
3,4 et 5 Octobre 2018

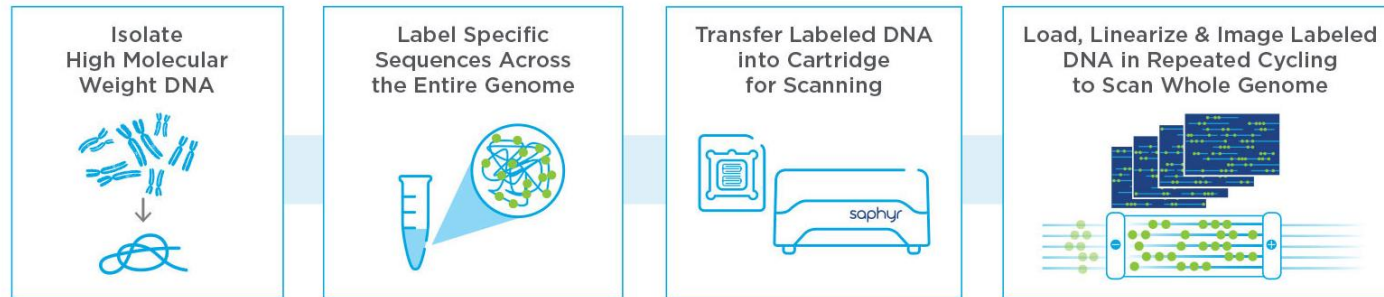
Faible poids moléculaire

HPM ~~~~~ Très HPM

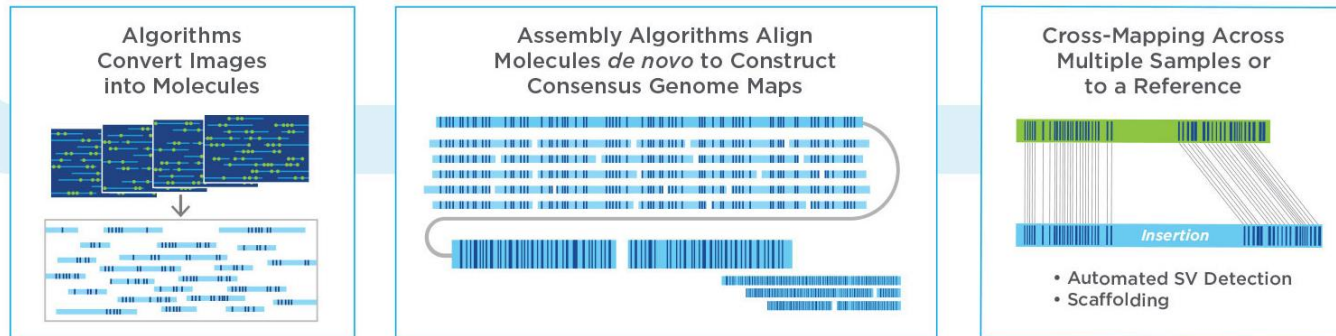




Technique de cartographie optique



High-throughput, High-resolution Imaging of Megabase Length Molecules

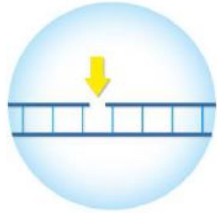


- Avantages: analyse de grandes molécules (150 kb à 2Mb), amélioration des assemblages, détection des SVs, haplotypage, étude de la réplication, étude des marques épigénétiques
- Inconvénients : préparation du matériel difficile, nécessite une référence de qualité.

MARQUAGES BIONANO

NLRS

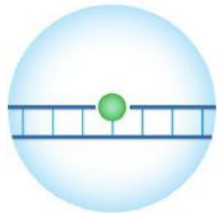
Nicking



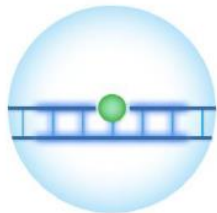
Labeling



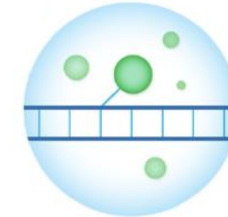
Repairing



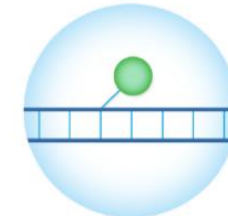
Staining



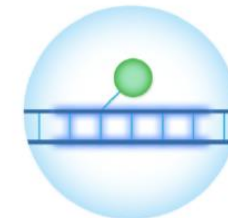
Labeling



Cleanup



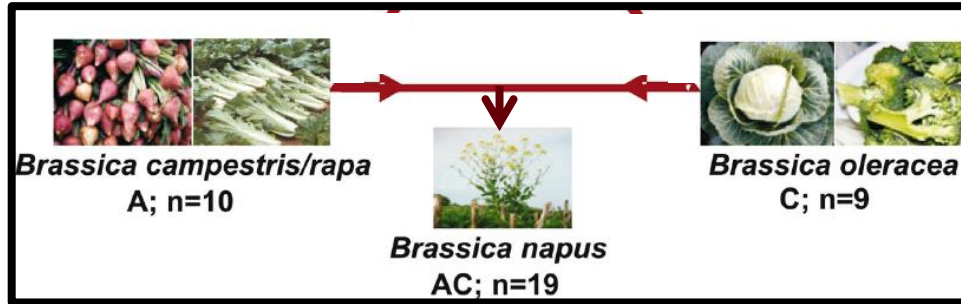
Homogenize
& Staining



DLS

Direct
Labeling
Stain

LES DIFFÉRENTS PROJETS



A diagram showing the evolutionary relationships between three Brassica species. On the left, *Brassica campestris/rapa* (A; n=10) is represented by images of carrots and corn. On the right, *Brassica oleracea* (C; n=9) is represented by images of a cabbage and broccoli. In the center, *Brassica napus* (AC; n=19) is represented by an image of a flowering plant. Red arrows indicate that *Brassica napus* is a hybrid of the other two species.

Brassica campestris/rapa
A; n=10

Brassica oleracea
C; n=9

Brassica napus
AC; n=19



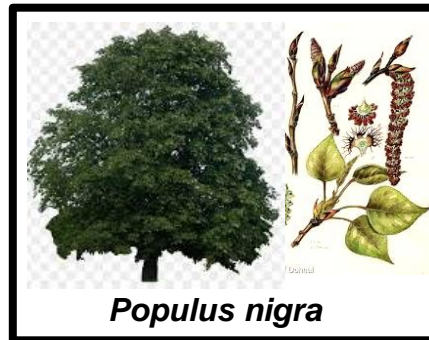
Two photographs of the parasitoid wasp *Cotesia glomerata*. The left image shows several wasps on a green leaf. The right image is a close-up of a cluster of white eggs attached to a leaf.

Cotesia glomerata



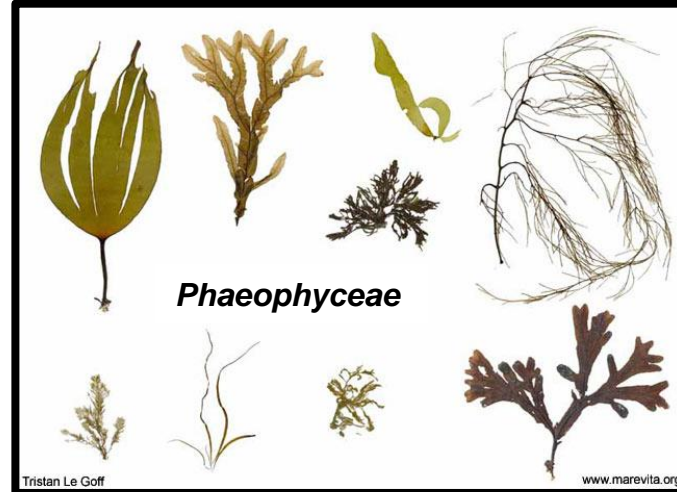
Two images of *Fagus sylvatica*: a large green tree and a smaller tree without leaves.

Fagus sylvatica



Two images of *Populus nigra*: a large green tree and a branch with catkins.

Populus nigra



A collection of various brown algae (Phaeophyceae) specimens, including leaves, stems, and root-like structures.

Phaeophyceae

Tristan Le Goff www.marevita.org



A photograph of the model plant *Arabidopsis thaliana*.

Arabidopsis thaliana



Two images of *Musa spp*: a banana plant in a pot and a bunch of yellow bananas.

Musa spp



A photograph of the agave-like plant *Espeletia schultzei* in a field.

Espeletia schultzei



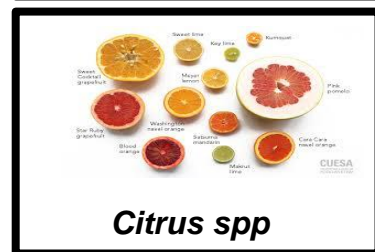
A photograph of several ripe, yellow-orange fruits of *Prunus spp* on a branch.

Prunus spp



A photograph of a clump of *Spartina* grass growing in a wet, sandy area.

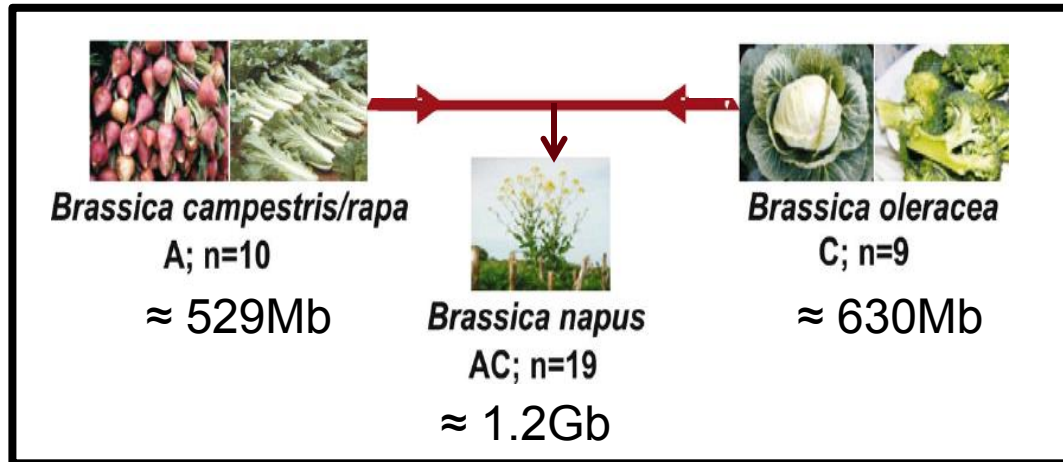
Spartina



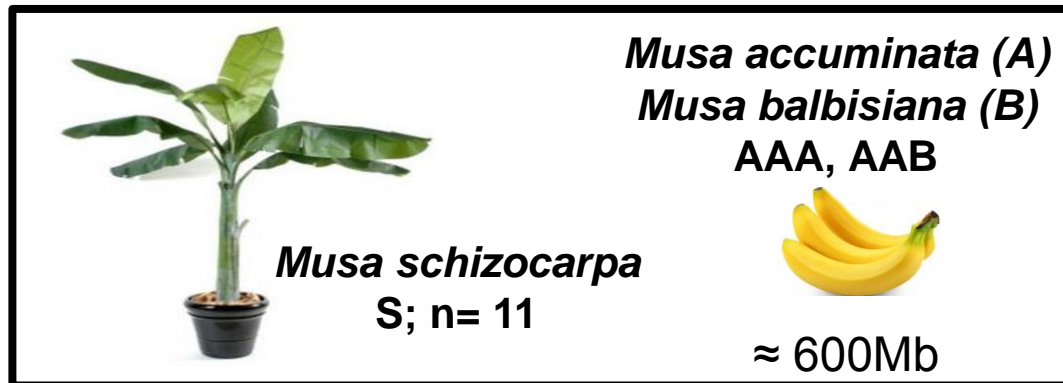
A collection of various citrus fruits, including oranges, lemons, and grapefruits, with labels for different varieties.

Citrus spp

CUESA



Human food
and oil production
Oilseed rape 3rd source of
vegetable oil in the world
and 1st in Europe



Human food
>117 millions t/year

Biotic and abiotic stresses: improvement of the selection
Structural variations, phasing: phylogeny, genetic diversity

Sequencing

Oxford Nanopore Technologies

ILLUMINA

Minlon



Promethlon



Optical mapping

BioNano Genomics

Saphyr



Wet lab

Plant samples

Very young leaves



Nuclei isolation & DNA extraction

Home-made protocols
Eva Hribova protocol
Bionano protocols



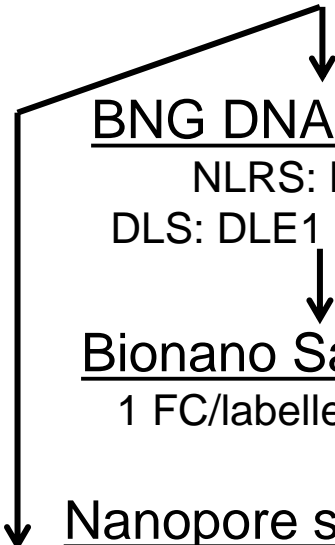
BNG DNA Labeling

NLRS: BspQI
DLS: DLE1 (beta-test)



Bionano Saphyr run

1 FC/labelled sample



Nanopore sequencing

DNA from plugs or from liquid extraction
Minlon or Promethlon

Bioinformatics

Nanopore reads

Assembly
Polishing



BNG molecules

de novo Assembly
2enzymes Hybrid scaffold



Final assembly

Polishing



High quality genome

Genes and TEs annotation

NUCLEI ISOLATION & DNA EXTRACTION

The most limiting step!!!!



Plants

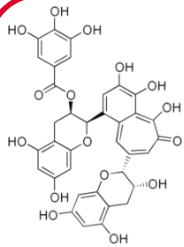


Young leaves

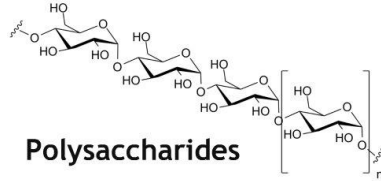


Dark treatment

Samples



Polyphenols

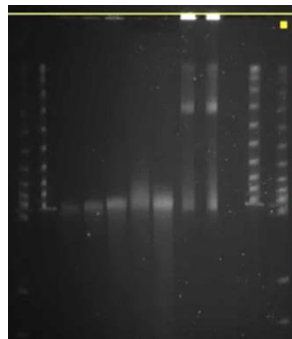


Polysaccharides



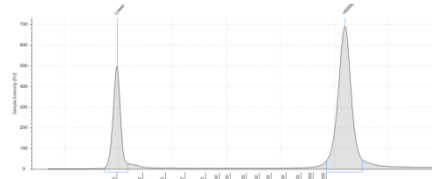
Isolate HMW DNA

BioNano protocols (4 for plants)
Home-made protocols

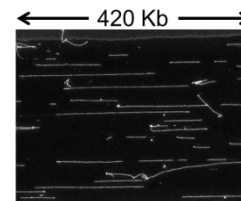


← 400kb

← 50kb



Argus® System/Opgen



HMW DNA Quality control

Tapestation (Agilent technologies)
Pippin Pulse (Sage Sciences)
Qcards Argus® System (Opgen)

For *Brassica spp.*:

Home-made protocol adapted from Zhang et al., 1995 and Zhang et al., 2012

Training of the collaborators (INRA, Rennes) at Genoscope

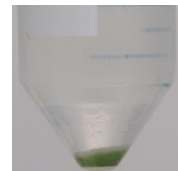
High quality DNA

For *Musa spp.*:

Tests on home-made protocols, bionano protocols did not succeed

Test Eva Hribova protocol adaptation: good for *M. schizocarpa* supernatant

g force	Genome size
1 200xg	> 6Gb
2 000xg	2-6Gb
2 500xg	0.3-2Gb
3 500xg	<0.3Gb



2500g



200g



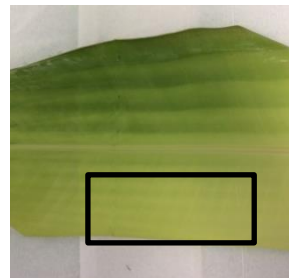
400g

Other species

M. schizocarpa



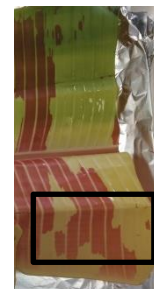
Good



Good (but...)



Not good



Not good

Brassica rapa et *Brassica oleracea*

+ *Brassica napus* 'synthétique' en cours

Brassica rapa : Z1 – yellow sarson – diploïde – 10 chr – ~529Mb

Brassica oleracea : HDEM – broccoli – diploïde – 9 chr – ~630Mb

Brassica napus 'synthétique' : RCC-S0 – tétraploïde – 19 chr – ~1,2Gb

Brassica rapa : Z1

Brassica oleracea : HDEM

Brassica napus 'synthétique' : RCC-S0



Séquençage et assemblage long-reads Nanopore
entre **32X** et **79X**



Constructions de cartes optiques Bionano
2 cartes : **BspQI** et **DLE-1**



Séquençage short-reads Illumina : polishing
paired-ends 250pb, PCR-free
environ **50X**



	<i>Brassica oleracea</i>	<i>Brassica rapa</i>
	HDEM	Z1
# Reads (>1Kb)	1,242,230	3,763,611
Cumulative size	20,081,668,921	31,496,053,430
Average size (bp)	16,165.8	8,368.57
N50 (bp)	32,339	15,042
N90 (bp)	7,718	3,570
Max size (bp)	607,633	582,421
# of reads > 50Kb	77,223	30,453
Coverage	31.87X	79X
Coverage (reads >50Kb)	8.23X	5.08X
Number of flowcells	14	12

	<i>Brassica oleracea</i>	<i>Brassica rapa</i>
	HDEM	Z1
N50 (bp)	32,339	15,042

	<i>Brassica oleracea</i>	<i>Brassica rapa</i>
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11

	<i>Brassica oleracea</i>	<i>Brassica rapa</i>
	HDEM	Z1
# of reads > 50Kb	77,223	30,453

	<i>Brassica oleracea</i>	<i>Brassica rapa</i>
	HDEM	Z1
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h

	<i>Brassica oleracea</i>	<i>Brassica rapa</i>
	HDEM	Z1
Number of <u>flowcells</u>	14	12

11h35

Assemblage de génomes - ONT panorama

Jean-Marc Aury– CEA-IbFJ/Génomscope, Evry

STATISTIQUES DES DONNÉES BRUTES BIONANO

	<i>B. rapa</i> Z1		<i>B. oleracea</i> HDEM	
	<u><i>BspQI</i></u>	DLE	<u><i>BspQI</i></u>	DLE
Total number of <u>molecules</u>	3,487,923	12,184,456	6,489,688	9,118,635
Total length <u>(Mbp)</u>	345,728	1,175,790	481,008	480,100
<u>Molecule N50</u> (kbp)	176	128	177.9	156.3
<u>Label density</u> (/100kb)	9.09	14.29	9.75	13.56

	<i>B. rapa</i> Z1		<i>B. oleracea</i> HDEM	
	<u>BspQI</u>	DLE	<u>BspQI</u>	DLE
<u>Number Genome Maps</u>	337	381	445	116
<u>Total Genome Map Length (Mbp)</u>	474.953	584.226	609.464	639.650
<u>Mean Genome Map Length (Mbp)</u>	1.409	1.533	1.370	5.514
<u>Median Genome Map Length (Mbp)</u>	0.838	0.464	0.845	0.606
<u>Genome Map N50 (Mbp)</u>	2.166	10.705	2.264	32.187

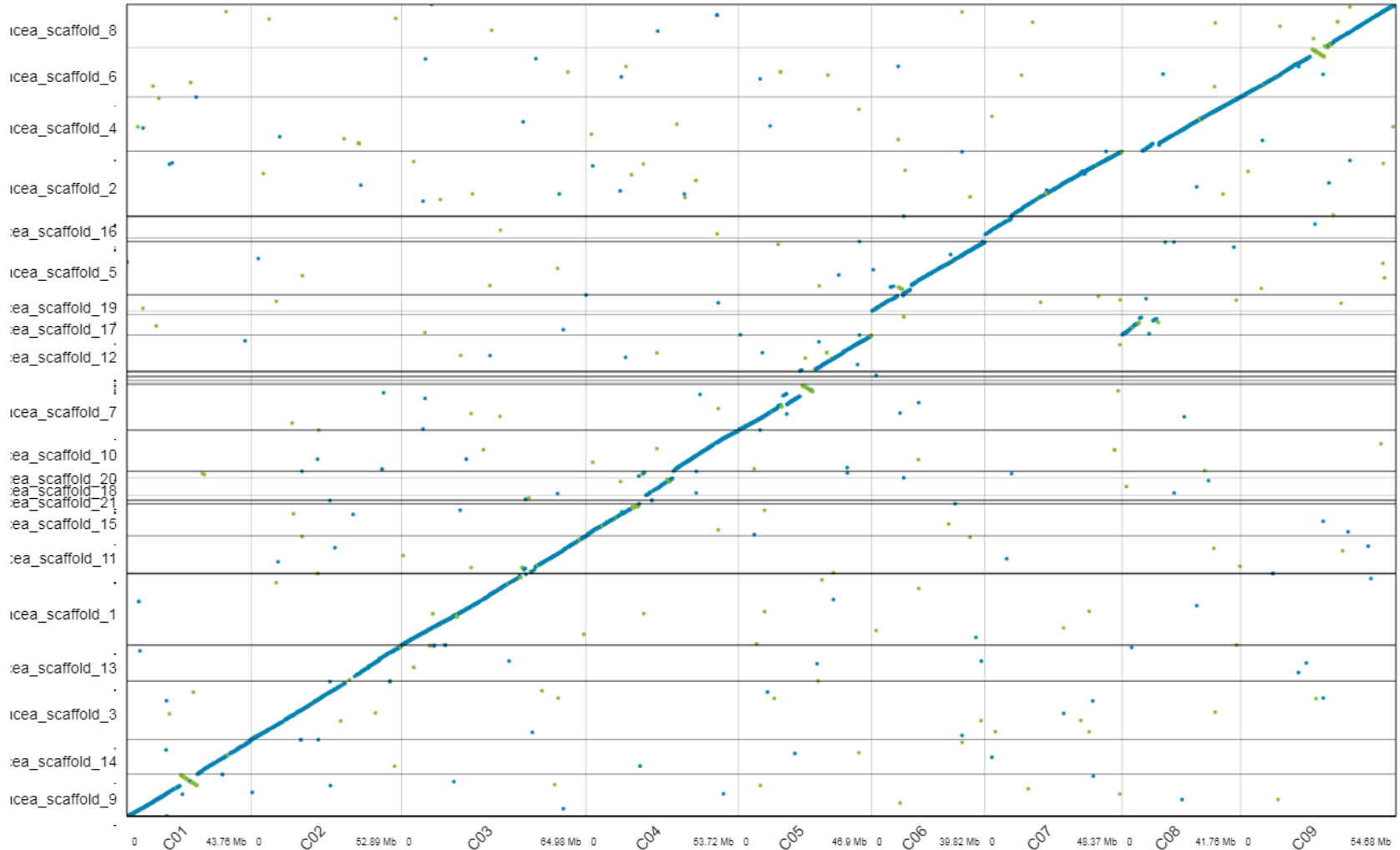
A rough assembly was 1st performed with parameters: -i 0 -V 0 -A -z -u -m (pipelineCL.py). This 1st result was used as a reference for a 2nd assembly, launched with the following parameters: -y -r (rough assembly cmap) -V 0 -m

Comparaison aux 1^{er} génomes de référence (TO1000 (2014) et Chiifu (2017) : short-reads assemblies)

	<i>Brassica oleracea</i>		<i>Brassica rapa</i>	
	To1000	HDEM	Chiifu	Z1
Reference	Liu et al. ¹⁵	This study MinION	Cai et al. ¹³	This study MinION
Estimated genome size	630	630	529	529
# scaffolds (≥2Kb)	1,428	140	86,852	335
Cumulative size	473,834,292	554,975,960	391,410,456	401,923,810
N50 (L50)	48,366,697 (5)	29,516,207 (8)	33,885,992 (5)	15,385,215 (8)
N90 (L90)	39,822,476 (9)	13,883,733 (17)	30,058 (212)	1,671,465 (31)
Max size	64,984,695	48,260,371	54,546,898	38,870,275
# of Ns	42,740,102 (9.02%)	9,958,104 (1.79%)	23,665,136 (6.04%)	32,963,474 (8.2%)

COMPARAISON GÉNOME TO1000 (SHORT-READS) VS HDEM (LONG- READS) : NUCMER

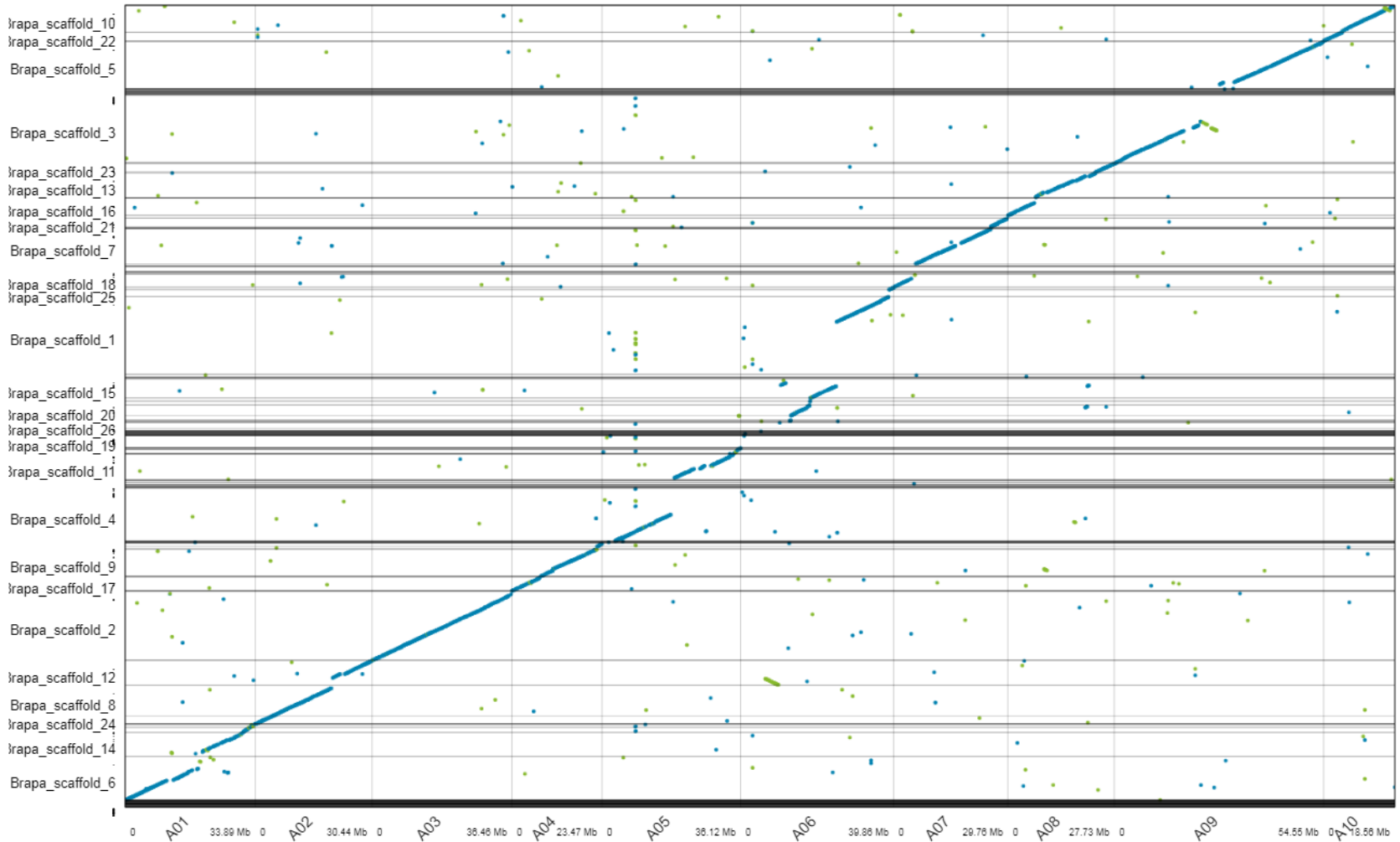
HDEM (long-reads) : 20 scaffolds les + longs



TO1000 (short-reads) : 9 pseudo-chromosomes

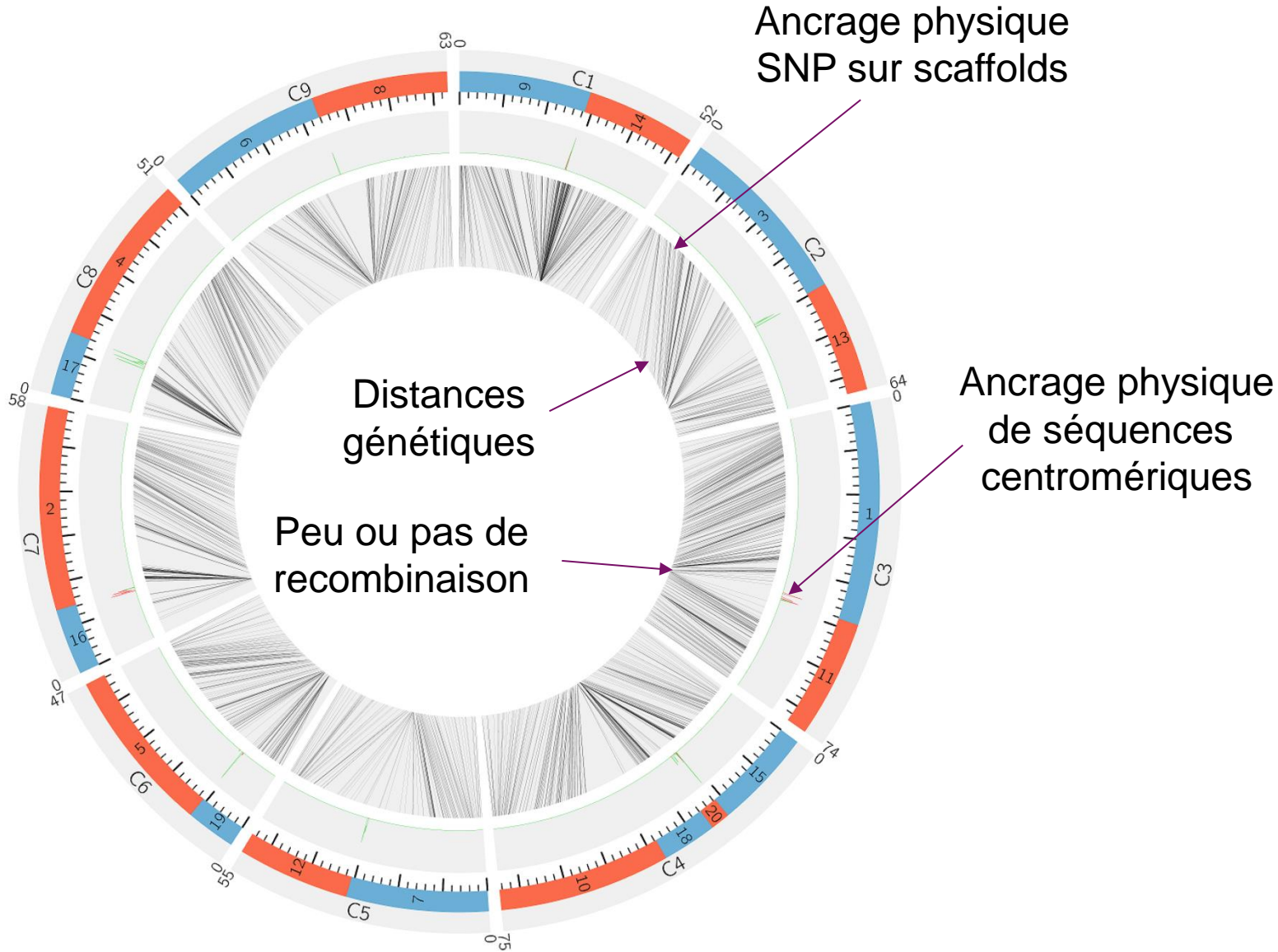
COMPARAISON GÉNOME CHIIFU (SHORT-READS) VS Z1 (LONG-READS) : NUCMER

Z1 (long-reads) : 26 scaffolds les + longs



Chiifu (short-reads) : 10 pseudo-chromosomes

COMPARAISON ASSEMBLAGE HDEM AVEC LA CARTE GÉNÉTIQUE RICHELAIN X HDEM



Genoscope: Patrick Wincker

LBioMEG: Valérie Barbe

Ghislaine Magdelenat

Barbara Mairey

Wahiba Berrabah

Sequencing Lab: Arnaud Lemainque

Thomas Guerin

Emilie Payen

Corinne Cruaud

Karine Labadie

R&D bioinformatics: Jean-Marc Aury

Caroline Belser

Benjamin Istace

Marion Dubarry

Benjamin Noel

Stefan Engelen

France Denoeud

INRA:

Biodiversity and Polyploidy team (part of Brassica group) : Anne-Marie Chèvre

Gwenaëlle Deniot

CIRAD:

Structure et évolution des génomes (Musa team): D'Hont Angélique

Franc-Christophe Baurens

Guillaume Martin

